

<u>Lieke J Ceton¹</u>, Marta G Montero¹, Fanny Grillet¹, Dieudonné J van der Meer¹, Willemijn Vader¹, Cor D de Kroon², Anne van Altena³, Nelleke Ottevanger³, Judith Kroep²

1. VitroScan B.V., Leiden, The Netherlands, 2. Leiden University Medical Center, 3. Radboud University Medical Center



Background

Immunotherapy brings great progress for cancers that are difficult to treat. It remains challenging to select sensitive patients upfront. PD-L1 expression, TMB and MSI/MSS status do not optimally differentiate between the sensitivity to individual immunotherapies.

We aim to improve patient stratification for immunotherapy by classification of patient-specific responses using a robust ex vivo 3D tumor testing platform

Material

Our study includes **101 patients** with predominantly ovarian carcinoma (80/101). Tumor tissue was collected in ongoing clinical trials from solids and liquids (Table 1). Five immune checkpoint inhibitors and a STING activator were tested.

Table 1: Sample characteristics						
Indication	#	Tissue type #				
Ovarian	80	Ascites	65			
		Solid tumor	15			
Bladder (B)	8	Solid tumor	8			
Lung (L)	4	Pleural fluid	4			
Breast	3	Pleural fluid	2			
		Biopsy	1			
Mesothelioma (M)	3	Pleural Fluid	3			
Melanoma	2	Cell line	2			
Cervix	1	Solid tumor	1			

Figure 1: Assay pipeline Drug exposure 3D Imaging 384 wells format Ex vivo tumor in native TME Drug effect on tumor morphology

w image Nuclei Tumoroids Overla

Method

Fresh tumor clusters were seeded into 384 well plates and exposed to different immunotherapies, while preserving the tumor micro-environment. In parallel, clusters were treated with chemo- and targeted therapies (not shown). Tumor killing and immune cell proliferation were measured using 3D image analysis (Figure 1). Sample and result quality are ensured through extensive automated quality control, combined with manual inspection by experts.

Ex vivo tumor sensitivity was classified as no response (<10%), weak (10-20%), strong (20-50%) and very strong (>50%), based on percentage tumor killing. Its statistical significance is expressed in two categories: significant (p-value: * < 0.05) and highly significant (** < 3.33e-4, Bonferroni corrected).

Results

Differential patient response profiles were observed (Main figure and Table 2).

For the current dataset, we measured a **highly significant immunotherapy response in ~35%** of the samples. **The assay has a technical success rate of 89%.**

Table 2:	All samples		SEA responders				
Immunotherapy response			Total		Highly significant response* (p < 0.00033)		
Treatments	Samples	Response	Samples	Response	Samples	Response	
SEA Superantigen	101	48.5 %	49	100 %	37	75.5 %	
CD3-CD28 beads	17	47.1 %	9	88.9 %	4	44.4 %	
Pembrolizumab	85	16.5 %	42	33.3 %	8	19.1 %	
Nivolumab	70	10.0 %	32	21.8 %	4	12.5 %	
Atezolizumab	64	12.5 %	34	23.5 %	6	17.7 %	
Ipilimumab	93	19.4 %	48	37.5 %	11	22.9 %	
Durvalumab	48	2.1 %	25	4.0 %	0	0 %	
ADU-S100	62	37.1 %	31	74.2 %	19	61.3 %	

Conclusion

This study reports the development of a robust ex vivo tumor testing platform that classified patient-specific sensitivity to 6 immunotherapies in over 100 patients, demonstrating the potential of ex vivo tumor testing to optimize patient stratification for immunotherapy.

Discussion

The platform enables better selection of patients in clinical trials for novel immunotherapies and in daily practice for those who could profit from immunotherapy, thereby avoiding toxicity in those who are not sensitive for these therapies. Clinical trials are ongoing to establish the correlation of our testing with **clinical** response to immunotherapy.



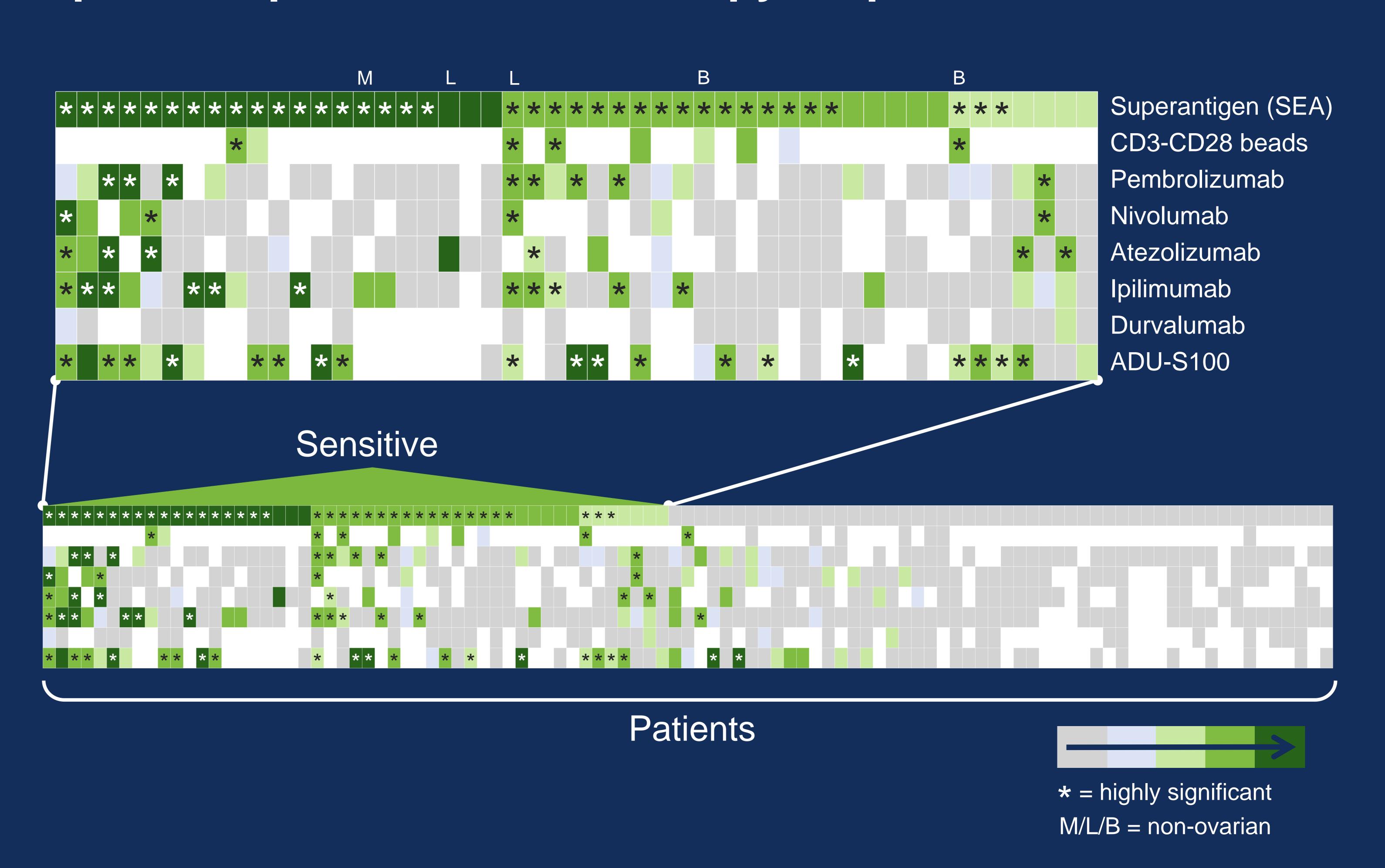








Ex-vivo 3D functional assay enables patient-specific immunotherapy response classification



Study of 100+ patients displays highly significant immunotherapy response profiles for 35% of the patients tested